

Result

Our wet lab experiment is consisted of three major parts, construction and testing of our circuit, testing of *mlrD* and *mlrA*, testing of riboswitches. Photos of electrophoresis¹ and results of sequencing will be listed as basic results of construction and results of HPLC and fluorescence measurement will work as efficiency verification of our project.

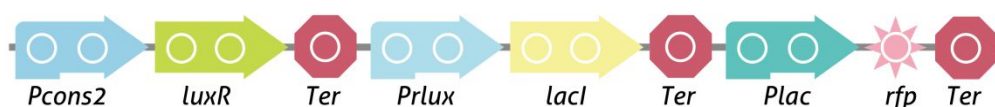
In actual experiment, our project is composed of three sections, the detector, the positive feedback circuit, the suicide circuit which are all used to test our design in experiment. In the process of construction, we first construct small devices from parts provided in the kits and then put all these devices together step by step.

Part 1:Circuit

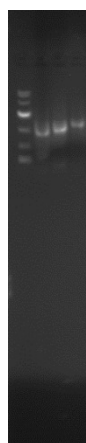
1. Detector circuit

The detector circuit is used to measure concentration of AHL and present fluorescence at certain concentration. It is made up of two plasmids.

(1)Plasmid 1: Pcons-rbs-luxR-T-Prlux-rbs-lacI-T-Plac-rbs-rfp-T



A. division 1 : Pcons2-rbs-luxR-T



Figure²: From left to right, Marker(DL10000), Pcons2, Pcons2-luxR, Pcons2-luxR-Ter³

Sequencing results:

We have ligated and sequenced this composite several times. Although the results of electrophoresis are identical to our expectation, the results of

¹ For specific procedures, please refer to the protocols.

² The actual photos of electrophoresis are listed in the attachment, while the results for each division maybe combination of results from different electrophoresis. However, we make sure that the marker matches while splicing.

³ Primers used in PCR are VF2 and VR, and following electrophoresis results are similar. .

sequencing always show that the where there is luxR is not it⁴. However, the same luxR in other composites meet our expectations. So, we suspect the authenticity of sequencing in this composite. We will verify luxR in our testing of function.

B. division 2: Prlux-rbs-lacI-T

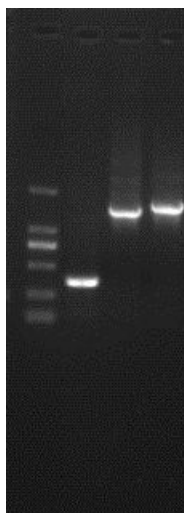


Figure ⁵: From left to right M: Marker (DL2000) , a: Prlux, Prlux-rbs-lacI, Prlux-rbs-lacI-T. The substances of electrophoresis are made from PCR of plasmids with VF2 and VR as their primers.

Sequencing result⁶:

```
CTATAAAATAGGCGTATCACGAGGCAGAATTCAGATAAAAAAATCCT
TAGCTTTTCGCTAAGGATGATTTCTGGAATTCGCGGCCGCTTCTAGAAAA
GAGGAGAAATTAAGCATGGTGAATGTGAAACCAGTAACGTTATACGATG
TCGCAGAGTATGCCGGTGTCTCTTATCAGACCGTTTCCCGCGTGGTGA
ACCAGGCCAGCCACGTTTCTGCGAAAACGCGGGAAAAAGTGGAAGCG
GCGATGGCGGAGCTGAATTACATTCCCAACCGCGTGGCACAACAACCTG
GCGGGCAAACAGTCGTTGCTGATTGGCGTTGCCACCTCCAGTCTGGC
CCTGCACGCGCCGTCGCAAATTGTCGCGGCGATTAAATCTCGCGCCGA
TCAACTGGGTGCCAGCGTGGTGGTGTGATGGTAGAACGAAGCGGCG
TCGAAGCCTGTAAAGCGGCGGTGCACAATCTTCTCGCGCAACGCGTC
AGTGGGCTGATCATTAACTATCCGCTGGATGACCAGGATGCCATTGCTG
TGGAAGCTGCCTGCACTAATGTTCCGGCGTTATTTCTTGATGTCTCTGA
CCAGACACCCATCAACAGTATTATTTTCTCCCATGAAGACGGTACGCGA
CTGGGCGTGGAGCATCTGGTTCGATTGGGTACACAGCAAATCGCGCT
GTTAGCGGGCCCATTAAGTTCTGTCTCGGCGCGTCTGCGTCTGGCTG
GCTGGCATAAATATCTCACTCGCAATCAAATTCAGCCGATAGCGGAACG
GGAAGGCGACTGGAGTGCCATGTCCGGTTTTCAACAAACCATGCAAAT
```

⁴ It is actually something there, however it doesn't match luxR.

⁵ To make the results clear, we get rid of band of other parts in this photo. The whole photo of electrophoresis will be listed later.

⁶ Primers used in sequence is VF2 and VR.

```
GCTGAATGAGGGCATCGTTCCCACTGCGATGCTGGTTGCCAACGATCA
GATGGCGCTGGGCGCAATGCGCGCCATTACCGAGTCCGGGCTGCGC
GTTGGTGCGGATATCTCGGTAGTGGGATACGACGATACCGAAGACAGC
TCATGTTATATCCCGCCGTTAACCACCATCAAACAGGATTTTCGCCTGC
TGGGGCAAACCAGCGTGGACCGCTTGCTGCAACTCTCTCAGGGCCAG
GCGGTGAAGGGCAATCAGCTGTTGCCCGTCTCACTGGTGAAAAGAAA
AACCACCCTGGCGCCCAATACGCAAACCGCCTCTCCCCGCGCGTTGG
CCGATTCATTAATGCAGCTGGCACGACAGGTTTCCCGACTGGAAAGCG
GGCAGGCTGCAAACGACGAAAACACTACGCTTTAGTAGCTTAATAACTCTG
ATAGTGCTAGTGTAGATCTCTACTAGTAGCGGCCGCTGCAGTCCGGCA
AAAAAGGGCAAGGTGTCACCACCCTGCCCTTTTTCTTTAAACCGAAA
AGATTACTTCGCGTTATGCAGGCTTCCTCGCTCACTGACTCGCTGCGC
TCGGTCGT
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C. Division 3: Plac-rbs-rfp-T

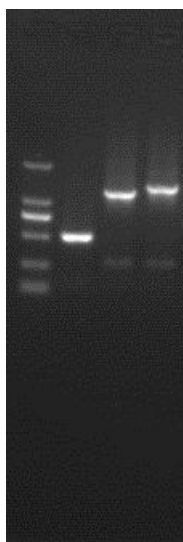
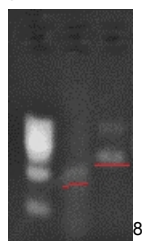


Figure : From left to right Marker (DL2000), Plac, Plac-rbs-rfp, Plac-rbs-rfp-T. The substances of electrophoresis are made from PCR of plasmid [s](#) with VF2 and VR as their primers.

Sequencing results:

The sequencing results is always wrong in this composite, but we actually saw the red fluorescence of colonies⁷. So, we stop the tedious job of sequencing and assume this composite is right.

D. Linking of Pcons2-rbs-luxR-T and Prlux-rbs-lacI-T



⁷ Shown in the following passages

⁸ For the reason of additional sequence up and down the aim sequence, the result line is a little longer.

Figure: From left to right, Marker(1Kb), Pcons2-rbs-luxR-Ter, Pcons2-rbs-luxR-Ter-Prlux-rbs-lacI-Ter
E. Linking of Pcons2-rbs-luxR-T-Prlux-rbs-lacI-T and Plac-rbs-rfp-T

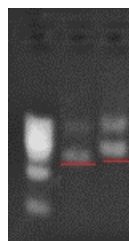
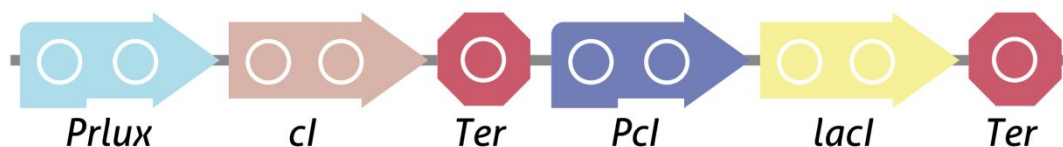


Figure: From left to right, Marker(1Kb), Pcons2-rbs-luxR-Ter-Prlux-rbs-lacI-Ter, Pcons2-rbs-luxR-Ter-Prlux-rbs-lacI-Ter-Plac-rbs-rfp-Ter

(2) Plasmid 2: Prlux-rbs-cl-T-Pcl-rbs-lacI-T



A. Division 1: Prlux-rbs-cl-T

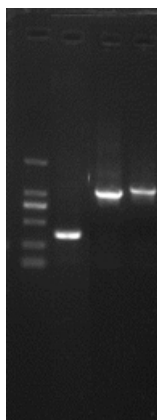


Figure: From left to right, Marker (DL2000) , Prlux, Prlux-rbs-cl, Prlux-rbs-cl-T. The substances of electrophoresis are made from PCR of plasmids with VF2 and VR as their primers.

B. Division 2: Pcl-rbs-lacI-T

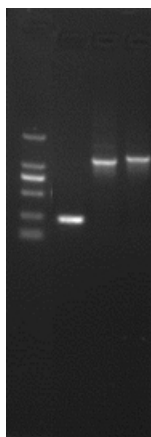


Figure: From left to right, Marker (DL2000) , Pcl, Pcl-rbs-lacI, Pcl-rbs-lacI-T. The substances of electrophoresis are made from PCR of plasmids with VF2 and VR as their primers.

C. Division 3: Prlux-rbs-cl-T-Pcl-rbs-lacI-T

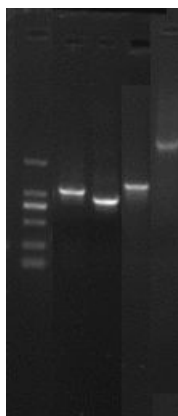


Figure: From left to right, Marker (DL2000) , Prlux-rbs-cl-T, Pcl-rbs-lacI-T, Prlux-rbs-cl-T-Pcl-rbs-lacI-T. The substances of electrophoresis are made from PCR of plasmids with VF2 and VR as their primers.

Sequencing results:

This part is rather long, and even one walking can't go through the whole sequence. So, we list three part of our sequence results.

VF2:

```
CCTATAAAATAGGCGTATCACGAGGCAGAATTTTCAGATAAAAA
AAATCCTTAGCTTTTCGCTAAGGATGATTTCTGGAATTCGCGGCCGC
TTCTAGAGACCTGTAGGATCGTACAGGTTTACGCAAGAAAATGGTT
TGTTATAGTCGAATAAATACTAGAAAAGAGGAGAAATTAAGCATGAG
CACAAAAAGAAACCATTAAACACAAGAGCAGCTTGAGGACGCACG
TCGCCTTAAAGCAATTTATGAAAAAAGAAAAATGAACTTGGCTTAT
CCCAGGAATCTGTGCGCAGACAAGATGGGGATGGGGCAGTCAGGC
GTTGGTGCTTTATTTAATGGCATCAATGCATTAAATGCTTATAACGC
CGCATTGCTTGCAAAAATTCTCAAAGTTAGCGTTGAAGAATTTAGC
CCTTCAATCGCCAGAGAAATCTACGAGATGTATGAAGCGGTTAGTA
TGCAGCCGTCACCTTAGAAGTGAGTATGAGTACCCTGTTTTTTCTCA
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TGTT CAGG CAGG GATGTTCTCACCTGAGCTTAGAACCTTTACCAAA
GGTGATGCGGAGAGATGGGTAAGCACAACCAAAAAAGCCAGTGAT
TCTGCATTCTGGCTTGAGGTTGAAGGTAATTCCATGACCGCACCAA
CAGGCTCCAAGCCAAGCTTTCTGACGGAATGTTAATTCTCGTTGA
CCCTGAGCAGGCTGTTGAGCCAGGTGATTTCTGCATAGCCAGACT
TGGGGGTGATGAGTTTACCTTCAAGAACTGATCAGGGATAGCGG
TCAGGTGTTTTTACAACCACTAAACCCACAGTACCCAATGATCCCA
TGCAATGAGAGTTGTTCCGTTGTGGGGAAAGTTATCGCTAGTCAGT
GGCCTGAAGAGACGTTTGGCGCTGCAaACGACgAAAACTACGCTT
TAGTAGCTT

VR:

ACGACCGAGCGCAGCGAGTCAGTGAGCGAGGAAGCCTGCAT
AACGCGAAGTAATCTTTTCGGTTTTAAAGAAAAAGGGCAGGGTG
TGACACCTTGCCCTTTTTTGGCGGACTGCAGCGGCCGCACTAGTG
AATTCTATAAACGCAGAAAGGCCACCCGAAGGTGAGCCAGTGTG
ACTCTAGTAGAGAGCGTTCACCGACAAACAACAGATAAAACGAAA
GGCCCAGTCTTTGACTGAGCCTTTGTTTTATTTGATGCCTGGCT
CTAGTAGAGATCTACACTAGCACTATCAGAGTTATTAAGCTACTAAA
GCGTAGTTTTCGTCGTTTGCAGCCTGCCCGCTTCCAGTCGGGAA
ACCTGTCGTGCCAGCTGCATTAATGAATCGGCCAACGCGCGGGGA
GAGGCGGTTTGCGTATTGGGCGCCAGGGTGGTTTTTCTTTTACC
AGTGAGACGGGCAACAGCTGATTGCCCTTCACCGCCTGGCCCTG
AGAGAGTTGCAGCAAGCGGTCCACGCTGGTTTGCCCCAGCAGGC
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C

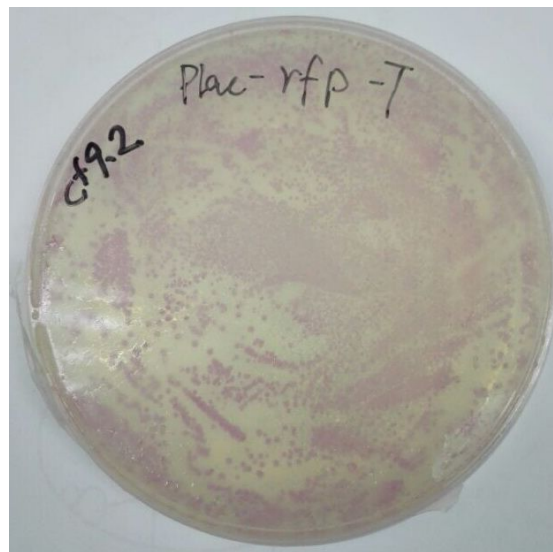
Walking(middle):

TTACCTTCAaGAaACTGATCAGGGATAGCGGTGAGGTGTTTTT
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TGTTCCGTTGTGGGGAAAGTTATCGCTAGTCAGTGGCCTGAAGAG
ACGTTTGGCGCTGCAAACGACGAAAACTACGCTTTAGTAGCTTAAT
AACGCTGATAGTGCTAGTGTAGATCGCTAATAAACTAGAGCCAGGC
ATCAAATAAAACGAAAGGCTCAGTCGAAAGACTGGGCCTTTGTTTT
TATCTGTTGTTTGTGCGGTGAACGCTCTCTACTAGAGTCACACTGGC
TCACCTTCGGGTGGGCCTTTCTGCGTTTATAGAATTCCTAGAGTA
ACACCGTGCGTGTTGACTATTTTACCTCTGGCGGTGATAATGGTTG
CTACTAGATGGTGAATGTGAAACAGTAACGTTATACGATGTGCGA
GAGTATGCCGGTGTCTCTTATCAGACCGTTTCCCGCGTGGTGAAC
CAGGCCAGCCACGTTTCTGCGAAAACGCGGGAAAAAGTGGAAGC
GGCGATGGCGGAGCTGAATTACATTCCTAACCGCGTGGCACAACA
ACTGGCGGGCAAACAGTCGTTGCTGATTGGCGTTGCCACCTCCA
GTCTGGCCCTGCACGCGCCGTGCGAAATTGTGCGGGCGATTAAAT
CTCGCGCCGATCAACTGGGTGCCAGCGTGGTGGTGTGCGATGGTA
GAACGAAGCGGCGTCGAAGCCTGTAAAGCGGCGGTGCACAATCT

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TCTCGCGCAACGCGTCAGTGGGCTGATCATTAACATCCGCTGGA
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TTTTCTCCCATGAAGACGGTACGCGACTGGGCGTGGAGCATCTGG
TCGCATTGGGTCACCAGCAAATCGCGCTGTTAGCGGGGCCATTAA
GTTCTGTCTCGGCGCGTCTGCGTCTGGCTGGCTGgCATAAATATCT
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GTT
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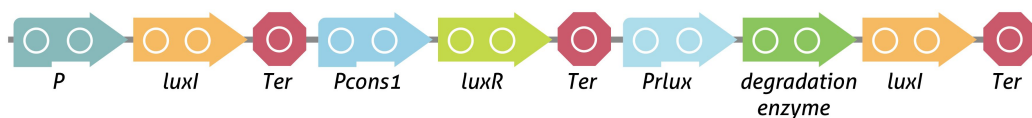
(3) Testing of function

A. The test of function of Plac-rfp-Ter shows that without lacI, rfp was expressed.



2. Positive feedback circuit.

(1) Construction of plasmid



Positive feedback system is consisted of four similar plasmids to test the efficiency of positive feedback and compare with results of modeling.

A. Division 1: Pcons2-rbs-lacI-T



Figure : From left to right, Marker (DL10000), Pcons2, Pcons2-lacI, Pcons-lacI-Ter.

Sequencing results:

```

CTATAAAAATAGGCGTATCACGAGGCAGAATTTTCAGATAAAAAAAT
CCTTAGCTTTTCGCTAAGGATGATTTCTGGAATTCGCGGCCGCTTCTAGA
GACCTGTAGGATCGTACAGGTTTACGCAAGAAAATGGTTTGTTATAGTC
GAATAAATACTAGAAAAGAGGAGAAATTAAGCATGGTGAATGTGAAACC
AGTAACGTTATACGATGTCTGCAGAGTATGCCGGTGTCTCTTATCAGACC
GTTTCCCGCGTGGTGAACCAGGCCAGCCACGTTTCTGCGAAAACGCG
GGAAAAAGTGAAGCGGCGATGGCGGAGCTGAATTACATTCCCAACC
GCGTGGCACAACAACCTGGCGGGCAAACAGTCGTTGCTGATTGGCGTT
GCCACCTCCAGTCTGGCCCTGCACGCGCCGTCGCAAATTGTCTCGCGGC
GATTAAATCTCGCGCCGATCAACTGGGTGCCAGCGTGGTGGTGTCTGAT
GGTAGAACGAAGCGGCGTCAAGCCTGTAAAGCGGCGGTGCACAATC
TTCTCGCGCAACGCGTCAGTGGGCTGATCATTAACTATCCGCTGGATG
ACCAGGATGCCATTGCTGTGGAAGCTGCCTGCACTAATGTTCCGGCGT
TATTTCTTGATGTCTCTGACCAGACACCCATCAACAGTATTATTTCTCC
CATGAAGACGGTACGCGACTGGGCGTGGAGCATCTGGTCTGCATTGGG
TCACCAGCAAATCGCGCTGTTAGCGGGCCCATTAAGTTCTGTCTCGGC
GCGTCTGCGTCTGGCTGGCTGGCATAAATATCTCACTCGCAATCAAATT
CAGCCGATAGCGGAACGGGAAGGCGACTGGAGTGCCATGTCCGGTTT
TCAACAAACCATGCAAATGCTGAATGAGGGCATCGTTCCCACTGCGAT
GCTGGTTGCCAACGATCAGATGGCGCTGGGCGCAATGCGCGCCATTA
CCGAGTCCGGGCTGCGCGTTGGTGCAGGATATCTCGGTAGTGGGATAC
GACGATACCGAAGACAGCTCATGTTATATCCCGCCGTTAACCACCATCA
AACAGGATTTTCGCCTGCTGGGGCAAACCAGCGTGGACCGCTTGCTG
CAACTCTCTCAGGGCCAGGCGGTGAAGGGCAATCAGCTGTTGCCCGT
CTCACTGGTGAAAAGAAAAACCACCCTGGCGCCCAATACGCAAACCGC
CTCTCCCCGCGCGTTGGCCGATTCATTAATGCAGCTGGCACGACAGGT
TTCCCGACTGGAAAGCGGGCAGGCTGCAAACGACGAAAACCTACGCTT
TAGTAGCTTAATAACTCTGATAGTGCTAGTGTAGATCTCTAATAAACTAGA
GCCAGGCATCAAATAAACGAAAGGCTCAGTCGAAAGACTGGGCCTTT
CGTTTTATCTGTTGTTTGTCTGGTGAACGCTCTCTACTAGAGTCACACTG
    
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GCTCACCTTCGGGTGGGCCTTTCTGCGTTTATAGAATTCAGTAGTGCG
GCCGCTGCAGTCCGGCAAAAAAGGGCAAGGTGTCACCACCCTGCCCT
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B. Division 2: Plac-rbs-luxI-T

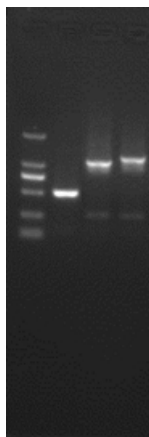


Figure: From left to right, Marker (DL2000), Plac, Plac-rbs-luxI, Plac-rbs-luxI-T. The substances of electrophoresis are made from PCR of plasmid with VF2 and VR as their primers.

Sequencing results:

ACCTATAaAAATAGGCGTATCACGAGGCAGAATTTTCAGATAAAAAA
ATCCTTAGCTTTTCGCTAAGGATGATTTCTGGAATTCGCGGCCGCTT
CTAGAGCAATACGCAAACCGCCTCTCCCCGCGCGTTGGCCGATTC
ATTAATGCAGCTGGCACGACAGGTTTCCCGACTGGAAAGCGGGCA
GTGAGCGCAACGCAATTAATGTGAGTTAGCTCACTCATTAGGCACC
CCAGGCTTTACACTTTATGCTTCCGGCTCGTATGTTGTGTGGAATT
GTGAGCGGATAACAATTTACACATACTAGAGAAAGAGGAGAAATT
AAGCATGACTATAATGATAAAAAAATCGGATTTTTTGGCAATTCCATC
GGAGGAGTATAAAGGTATTCTAAGTCTTCGTTATCAAGTGTTTAAGC
AAAGACTTGAGTGGGACTTAGTTGTAGAAAATAACCTTGAATCAGA
TGAGTATGATAACTCAAATGCAGAATATATTTATGCTTGTGATGATAC
TGAAAATGTAAGTGGATGCTGGCGTTTATTACCTACAACAGGTGATT
ATATGCTGAAAAGTGTTTTTCCTGAATTGCTTGGTCAACAGAGTGC
TCCCAAAGATCCTAATATAGTCGAATTAAGTCGTTTTGCTGTAGGTA
AAAATAGCTCAAAGATAAATACTCTGCTAGTGAAATTACAATGAAA
CTATTTGAAGCTATATATAAACACGCTGTTAGTCAAGGTATTACAGAA
TATGTAAACAGTAACATCAACAGCAATAGAGCGATTTTTAAAGCGTAT
TAAAGTTCCTTGTCATCGTATTGGAGACAAAGAAATTCATGTATTAG
GTGATACTAAATCGGTTGTATTGTCTATGCCTATTAATGAACAGTTTA
AAAAAGCAGTCTTAAATGCTGCAAACGACGAAAACACTACGCTTTAGT
AGCTTAATAACTCTGATAGTGCTAGTGTAGATCTCTAATAATACTAGA
GCCAGGCATCAAATAAAACGAAAGGCTCAGTCGAAAGACTGGGCC
TTTCGTTTTATCTGTTGTTTGTGCGGTGAACGCTCTCCACTCGGTCA
CACTGAGAACTAGTAGCGGCCGCTGCAGTCCGGCAAAAAAGGGC

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AAGGTGTCACCACCCTGCCCTTTTCTTTAAACCGAAAAGATTAC
TTCGCGTTATGCAGGCTTCCTCGCTCACTGACTCGCTGCGCTCGG
TCGTTTCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAA
TACGGTTATCCACAGAATCAGGGGATAACGCAGGAAAGAACATGT
GAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCG
TTGCTGGCGTTTTTCCACAGGCTCCGCCCCCTGACGAGCATCAC
AAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAgGACTA
TAAAGATACCAGGCGTTTCCCCCTGGAAGCTCCCTCGTGCGCTCT
CCTGTTCCGACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCTC
CCTTCGGGAAGCGT
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C. Division 3: Pcons-rbs-luxR-T

The figure is same with the one in the Detector circuit division 1.

Sequencing results:

The same with that in the Detector circuit division 1.

D. Division 4: Prlux-rbs-rfp-T, Prlux-rfp-luxI-T, Prlux-rfp-luxR-T, Prlux-rfp-luxI-luxR-T

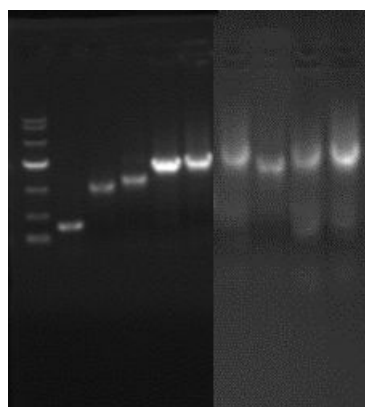


Figure: From left to right, Marker(DL10000), Prlux, Prlux-rbs-rfp, Prlux-rbs-rfp-Ter, Prlux-rbs-rfp-rbs-luxI, Prlux-rbs-rfp-rbs-luxI-Ter, Prlux-rbs-rfp-rbs-luxR-Ter, Prlux-rbs-rfp-luxR, Prlux-rbs-rfp-luxI-rbs-luxR, Prlux-rbs-rfp-rbs-luxI-rbs-luxR-Ter.

Sequencing results:

Prlux-rbs-rfp-T:

```
TATAAaAATAGGCGTATCACGAGGCAGAAATTCAGATAAAAAAA
ATCCTTAGCTTTTCGCTAAGGATGATTTCTGGAATTCGCGGCCGCTT
CTAGAGACCTGTAGGATCGTACAGGTTTACGCAAGAAAATGGTTTG
TTATAGTCAATAAATACTAGAAAAGAGGAGAAAATGGTGAGCAAG
GGCGAGGAGGATAACATGGCCATCATCAAGGAGTTTCATGCGCTTC
AAGGTGCACATGGAGGGCTCCGTGAACGGCCACGAGTTCGAGAT
CGAGGGCGAGGGCGAGGGCCGCCCTACGAGGGGCACCCAGACC
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CGAGGGCTTCAAGTGGGAGCGCGTGATGAACTTCGAGGACGGCG
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GGCCCCGTAATGCAGAAGAAGACCATGGGCTGGGAGGCCTCCTC
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AGCAGAGGCTGAAGCTGAAGGACGGCGGCCACTACGACGCTGAG
GTCAAGACCACCTACAAGGCCAAGAAGCCCGTGACGCTGCCCGG
CGCCTACAACGTCAACATCAAGTTGGACATCACCTCCCACAACGA
GGACTACACCATCGTGGAACAGTACGAACGCGCCGAGGGGCCGCC
ACTCCACCGGCGGCATGGACGAGCTGTACAAGTAATAATACTAGA
GCCAGGCATCAAATAAAACAAAAGGCTCAGTCGAAAGACTGGGCC
TTTCGTTTTATCTGTTGTTTGTGCGGTGAACGCTCTCTACTAGAGTCA
CACTGGTTCACCTTCGGGTGGGCCTTTCTGCGTTTATAGAATTAC
TAGTGCGGCCGCTGCAGTCCGGCAAAAAGGGCAAGGTGTCACC
ACCCTGCCCTTTTTCTTTAAACCGAAAAGATTACTTCGCGTTATGC
AGGCTTCCTCGCTCACTGACTCGCTGCGCTCG_gTCGT

Prlux-rbs-rfp-rbs-luxl-T:

aTTAACCTATAAAAATAGGCGTATCACGAGGCAGAAATTTTCAGAT
AAAAAAATCCTTAGCTTTTCGCTAAGGATGATTTCTGGAATTCGCG
GCCGCTTCTAGAGACCTGTAGGATCGTACAGGTTTACGCAAGAAA
ATGGTTTGTATAGTCGAATAAATACTAGAAAAGAGGAGAAAATGGT
GAGCAAGGGCGAGGAGGATAACATGGCCATCATCAAGGAGTTCAT
GCGCTTCAAGGTGCACATGGAGGGCTCCGTGAACGGCCACGAGT
TCGAGATCGAGGGCGAGGGCGAGGGCCGCCCTACGAGGGCAC
CCAGACCGCCAAGCTGAAGGTGACCAAGGGTGGCCCCCTGCCCT
TCGCCTGGGACATCCTGTCCCCCTCAGTTCATGTACGGCTCCAAGG
CCTACGTGAAGCACCCCGCCGACATCCCCGACTACTTGAAGCTGT
CCTTCCCCGAGGGCTTCAAGTGGGAGCGCGTGATGAACTTCGAG
GACGGCGGCGTGGTGACCGTGACCCAGGACTCCTCCTTGACGGA
CGGCGAGTTCATCTACAAGGTGAAGCTGCGCGGCACCAACTTCC
CCTCCGACGGCCCCGTAATGCAGAAGAAGACCATGGGCTGGGAG
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GCTGCCCGGCGCCTACAACGTCAACATCAAGTTGGACATCACCTC
CCACAACGAGGACTACACCATCGTGGAACAGTACGAACGCGCCG
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CCTGAATTGCTTGGTCAACAGAGTGCTCCCAAAGATCCTAATATAG
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AACTCTGCTAGTGAAATTACAATGAACTATTTGAAGCTATATATAAA
CACGCTGTTAGTCAAGGTATTACAGAATATGTAACAGTAACATCAAC
AGCAATAGAGCGATTTTTAAAGCGTATTAAAGTTCCTTGTCATCGTA
TTGGAGACAAAGAAATTCATGTATTAGGTGATACTAAATCGGTTGTA
TTGTCTATGCCTATTAATGAACAGTTTAAAAAAGCAGTCTTAAATGC
TGCAAACGACGAAAACACTACGCTTTAGTAGCTTAATAACTCTGATAGT
GCTAGTGTAGATCTCTAATAATACTAGAGCCAGGCATCAAATAAAAC
GAAAGGCTCAGTCGAAAGACTGGGCCTTTCGTTTTATCTGTTGTTT
GTCGGTGAACGCTCTCCACTCGGTCACTGAGAACTAGTAGCGG
CCGCTGCAGTCCGGCAAAAAGGGCAAGGTGTCACCACCCTGCC
CTTTTTCTTTAAACCGAAAAGATTACTTCGCGTTATGCAGGCTTCC
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Prlux-rbs-rfp-rbs-luxR-T:

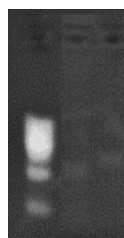
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Prlux-rbs-rfp-rbs-luxI-rbs-luxR-Ter

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```

E. Linking of Pcons2-rbs-lacI-T and Plac-rbs-luxI-T



From left to right, Marker(1Kb), Pcons2-rbs-lacI-Ter, Pcons2-rbs-lacI-Ter-Plac-rbs-luxI-Ter

Sequencing results:

```
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F. Linking of Pcons2-rbs-lacI-T-Plac-rbs-luxI-T and Pcons2-rbs-luxR-T

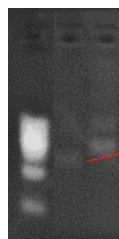


Figure: From left to right, Marker(1Kb), Pcons2-rbs-lacI-T-Plac-rbs-luxI-T, Pcons2-rbs-lacI-T-Plac-rbs-luxI-T-Pcons2-rbs-luxR-Ter.

G. Completion of four systems.

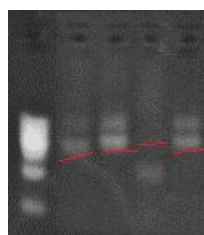
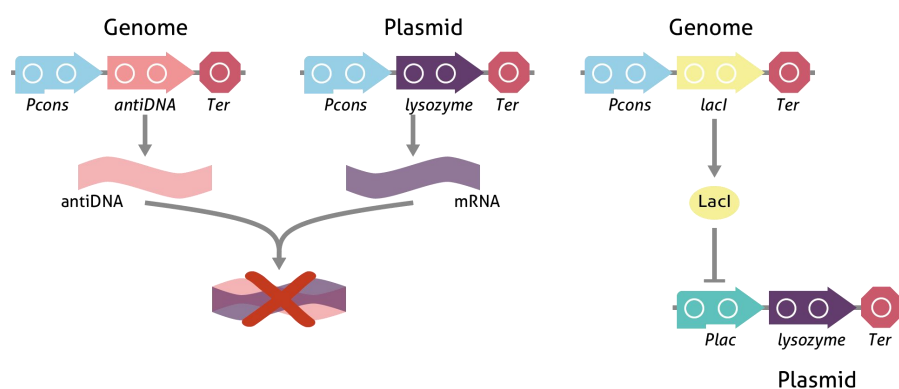


Figure: From left to right, Marker(1Kb), Pcons2-rbs-lacI-T-Plac-rbs-luxI-T-Pcons2-rbs-luxR-Ter, Pcons2-rbs-lacI-T-Plac-rbs-luxI-T-Pcons2-rbs-luxR-Ter-Prlux-rbs-rfp-Ter, Pcons2-rbs-lacI-T-Plac-rbs-luxI-T-Pcons2-rbs-luxR-Ter-Prlux-rbs-rfp-rbs-luxI-Ter, Pcons2-rbs-lacI-T-Plac-rbs-luxI-T-Pcons2-rbs-luxR-Ter-Prlux-rbs-rfp-rbs-luxI-rbs-luxR-Ter.

(2) Testing of function

3. Suicide circuit



(1) Construction of plasmid

A. Division 1 PgoITS-golS-PgolB-rbs-tetR-Ter

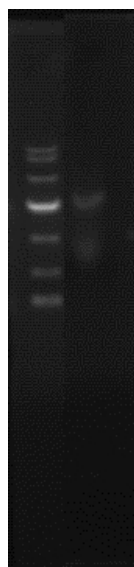


Figure: From left to right, Marker(DL10000), PgolTS-golS-PgolB-rbs-tetR-Ter

Sequencing results:

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B. Division 2 Ptet-rbs-rfp-T:

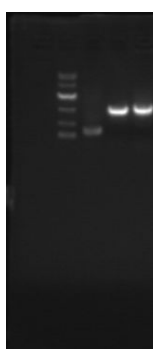


Figure: From left to right, Marker (DL2000), Ptet, Ptet-rbs-rfp, Ptet-rbs-rfp-T. The substances of electrophoresis are made from PCR of plasmids with VF2 and VR as their primers.

Sequencing results:

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C. Division 3 Plac-rbs-tetR-T:

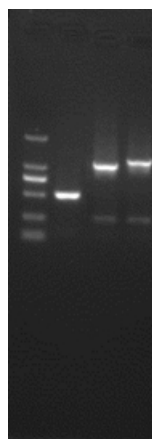


Figure: From left to right, Marker (DL2000) , Plac, Plac-rbs-tetR, Plac-rbs-tetR-T. The substances of electrophoresis are made from PCR of plasmid with VF2 and VR as their primers.

Sequencing results:

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CTACGGCTACCTAGAAGAACAGTAAT
```

D. Division 4 Pcons-rbs-lacI-T:

The figure is the same with that in the positive feedback circuit.

Sequencing results:

It is the same with that in the positive feedback circuit.

E. Linking of P_{golTS}-golS-P_{golB}-rbs-tetR-Ter and P_{tet}-rbs-rfp-Ter

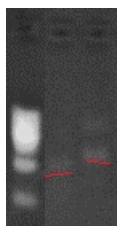


Figure: From left to right, Marker(1Kb), P_{golTS}-golB-P_{golB}-tetR-Ter, P_{golTS}-golB-P_{golB}-tetR-Ter-P_{tet}-rbs-rfp-Ter.

F. Linking of P_{golTS}-golS-P_{golB}-rbs-tetR-Ter-P_{tet}-rbs-rfp-T and Plac-rbs-tetR-T

G. Linking of P_{golTS}-golS-P_{golB}-rbs-tetR-Ter-P_{tet}-rbs-rfp-T-Plac-rbs-tetR-T and Pcons2-rbs-lacI-T

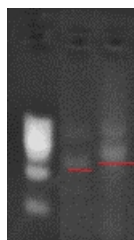


Figure: From left to right, Marker(1Kb), P_{golTS}-golB-P_{golB}-tetR-Ter-P_{tet}-rbs-rfp-Ter, P_{golTS}-golB-P_{golB}-tetR-Ter-P_{tet}-rbs-rfp-Ter-Plac-rbs-tetR-Ter-Pcons2-rbs-lacI-Ter.

(2) Testing of function

A. The test of function of P_{tet}-rbs-rfp-Ter shows that without tetR, rfp was expressed.



The left one is Ptet-rbs-rfp-Ter, and the right one is another plasmid. The photo shows that without tetR, rfp will be produced, and bacterial will get red.

4. OMPA-golB:



Figure: From left to right, Marker(DL10000), OMPA-golB.

Sequencing results:

```
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ACCAGGTTAACCCGTATGTTGGCTTTGAAATGGGTTACGACTGGTTAGGTC
GTATGCCGTACAAAGGCAGCGTTGAAAACGGTGCATACAAAGCTCAGGGC
GTTCAACTGACCGCTAAACTGGGTTACCCAATCACTGACGACCTGGACATC
TACACTCGTCTGGGTGGCATGGTATGGCGTGCAGACACTAAATCCAACGTT
TATGGTAAAAACCACGACACCGGCGTTTCTCCGGTCTTCGCTGGCGGTGTT
GAGTACGCGATCACTCCTGAAATCGCTACCCGTCTGGAATACCAAGTGGACC
AACACATCGGTGACGCACACACCATCGGCACTCGTCCGGACAACGGAGT
CGACATGCAGTTCCATATTGATGACATGACCTGCGGCGGCTGCGCCAGTAC
GGTAAAAAAGACGATTCTGACTCTCGATGCTAATGCGACGGTGAGAACTGA
CCCGGCGACGCGTCTGGTTGACGTTGAAACGTCGCTATCCGCGGAGCAGA
TTGCCGCCGCCCTGCAAAAGGCCGTTTCCCGCCGCGCGAGAGGCTCGA
GGATTACAAGGATGACGACGATAAGATCTAATAATACTAGTAGCGGCCGCTG
CAGTCCGGCAAAAAAGGGCAAGGTGTCACCACCCTGCCCTTTTCTTTAAA
ACCGAAAAGATTACTTCGCGTTATGCAGGCTTCCTCGCTCACTGACTCGCT
GCGCTCGTC
```

5. MlrD and MrlA

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(1) Construction of plasmid

MlrD:



Figure: From left to right, Marker(DL10000), MlrD.

Sequencing result:

```
GATCaACCAGCGGTCTTACTGCTGGCTGCGACGGAAATGTGGGAGGC
CTTTTCCTATGTCGGGCTCAGAACCGTACTGGTCTACTACCTTACGCA
GGACCTGGGCTATTCGACCGAGGACGCCTCACTTATCTATGGGACGT
TCCTCGGCGTAGCCTATGTAACGCCAATCCTCGGTGGGTGGATCGCC
GATAGGTTTATTGGCCGTTTCGGCGGCAATTGTCGGTGGCGCATTGCT
GAAGATGGCCGGGTACATCGGCCTTTTGCTTGGCGCGAACGTTACGG
GCTGCCTCGCCGCAATTGTCATTGGCAATGGCCTGTTTCTTCCAACCTC
TGCCCGCTACGCTGGGTGCTCTTTTCTCGCCGAACGACCCCGATCGC
CAGCGCAGTTTCAGCTTCTACTATCTCGCAGTGAGCGCTGGTGGCT
GCTGGCACCGCTGATCTGCGGCACGCTTGGAGAGAATTCGGCTGG
CGGTACAGTTTCCTCGCTTCCGCTACCGGACTTGCAGCAGCCATCGT
TATCTTTCTCGCCGGACGCCATCTTCTGCCTCCAGACCGACCTGGAG
CAGCGTCCCGTCTGGTCGACGAAACGCCGGTCGCGCAGGCGAGCC
AGTCCGTCATCCCGCTCCTGGCAGGTGTCCTTGCAGCAGTAATCGTC
TTGCGGGTCGCTTATGAGCAACTCGGCAACACTGTCGCGCTTTTCGC
CGCCAGCCAGGTGATCGTTGCTAGGGGCAGATATCACCATCCCTT
ATACCTGGTTTCAGTCGCTCAATCCGCTGGGGGTCAATTCTGTTACCCC
CGCTCCTCGTCTGGGGCTGGCGTAAGGCCGCCGCGAGAGGTGGCT
CGCAGAACGACTATTTAGGATGGCCATTGGTAGCTGCATCATGGCC
GCGGCCTTTGTTGGGCTCGCTCTGATCATCCATCTCGGGCAACCCGG
TCAGATTTTCTGGCCTGTTCTAGCAGCGTTTTTTCTGAAGTAACCTTT
GGCGAGTTGTGGGTGCTTCCGGTCGGTCTCAGTCTGTTTGCGCGCTT
GGCACCCGCAGGGCGAGGTGCAGTCACCATCGCGTTCTGGTATAGC
GCGCGAGCCGCTGGCAATTTTCTTGCCGGATTGATGGGGCGCGCCG
AACCTGCACTCGGATATGGCAACTTCTTCCTGCTTTGCGCAGTGTTC
CGCTGCTGGCAGCAACGATCTTCGTCGCGATCGGCAAGCGCTCGC
```

(2) Testing of function

6. Attachment

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(1) Complete photos of electrophoresis

A.

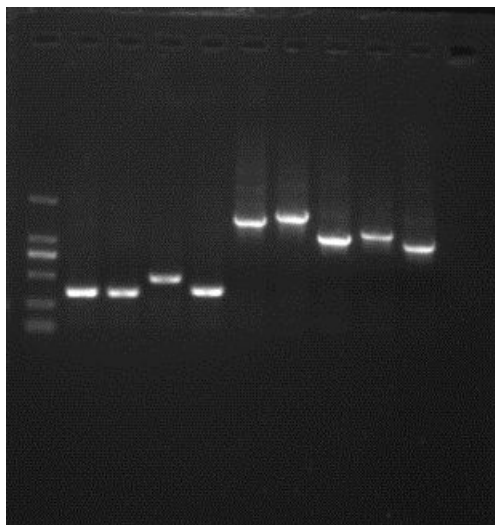


Figure: From left to right, Marker(DL2000), Pcons, Pcons-rbs-luxR, Pcons-rbs-luxR-T, Prlux, Prlux-rbs-lacI, Prlux-rbs-lacI-T, Prlux-rbs-CI, Prlux-rbs-CI-T, Prlux-rbs-rfp. The substances of electrophoresis are made from PCR of plasmids with VF2 and VR as their primers.

B.

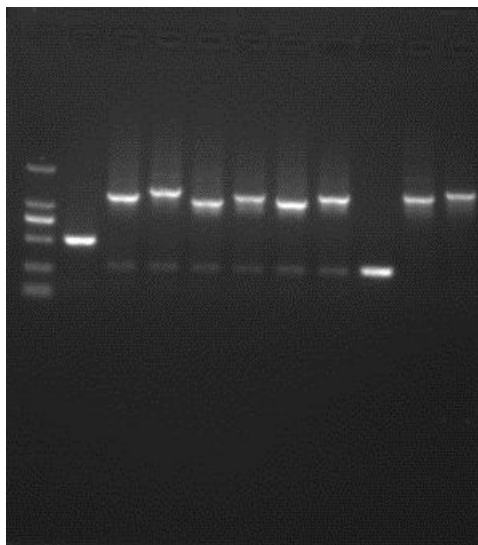


Figure: From left to right, Marker(DL2000), Plac, Plac-rbs-rfp, Plac-rbs-rfp-T, Plac-rbs-luxI, Plac-rbs-luxI-T, Plac-rbs-tetR, Plac-rbs-tetR-T, PCI, PCI-rbs-lacI, PCI-rbs-lacI-T

. The substances of electrophoresis are made from PCR of plasmid [s](#) with VF2 and VR as their primers.

C.

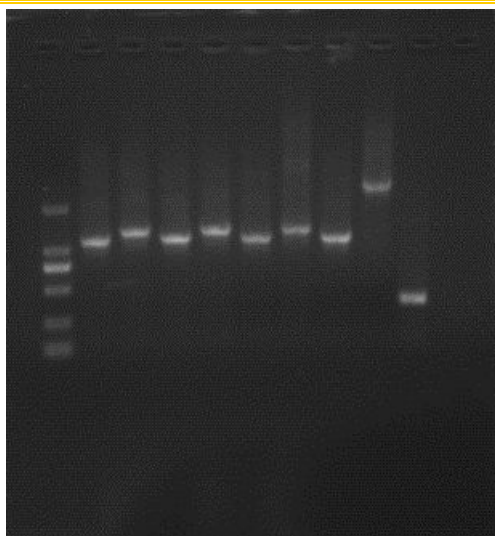


Figure: From left to right, Marker(DL2000) Prlux-rbs-rfp, Prlux-rbs-rfp-T, Prlux-rbs-rfp-rbs-luxI, Prlux-rbs-rfp-rbs-luxI-T, Prlux-rbs-rfp-rbs-luxR, Prlux-rbs-rfp-rbs-luxR-T, Prlux-rbs-rfp-rbs-luxI-rbs-luxR, Prlux-rbs-CI-T-PCI-rbs-lacI-T, Ptet. The substances of electrophoresis are made from PCR of plasmids with VF2 and VR as their primers.

D.

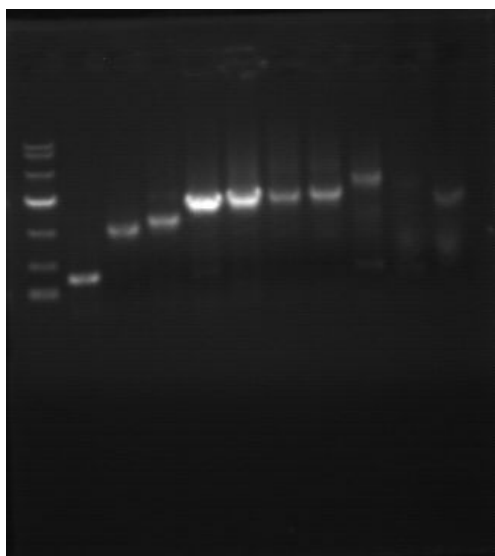


Figure: From left to right, Marker(DL10000), Prlux, Prlux-rbs-rfp, Prlux-rbs-rfp-Ter, Prlux-rbs-rfp-luxI, Prlux-rbs-rfp-rbs-luxI-Ter, Prlux-rbs-rfp-rbs-luxR, Prlux-rbs-rfp-rbs-luxR-Ter, Prlux-rbs-rfp-rbs-luxI-rbs-luxR, Prlux-rbs-rfp-rbs-luxI-rbs-luxR-Ter, PgoITS-golS-PgolB-rbs-tetR-Ter.

E.

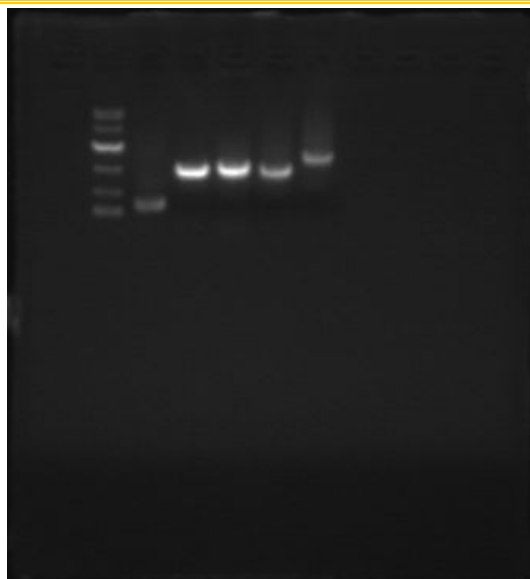


Figure: From left to right, Marker(DL10000), Ptet, Ptet-rbs-rfp, Ptet-rbs-rfp-Ter, OMPA-golB, MirD.

F.

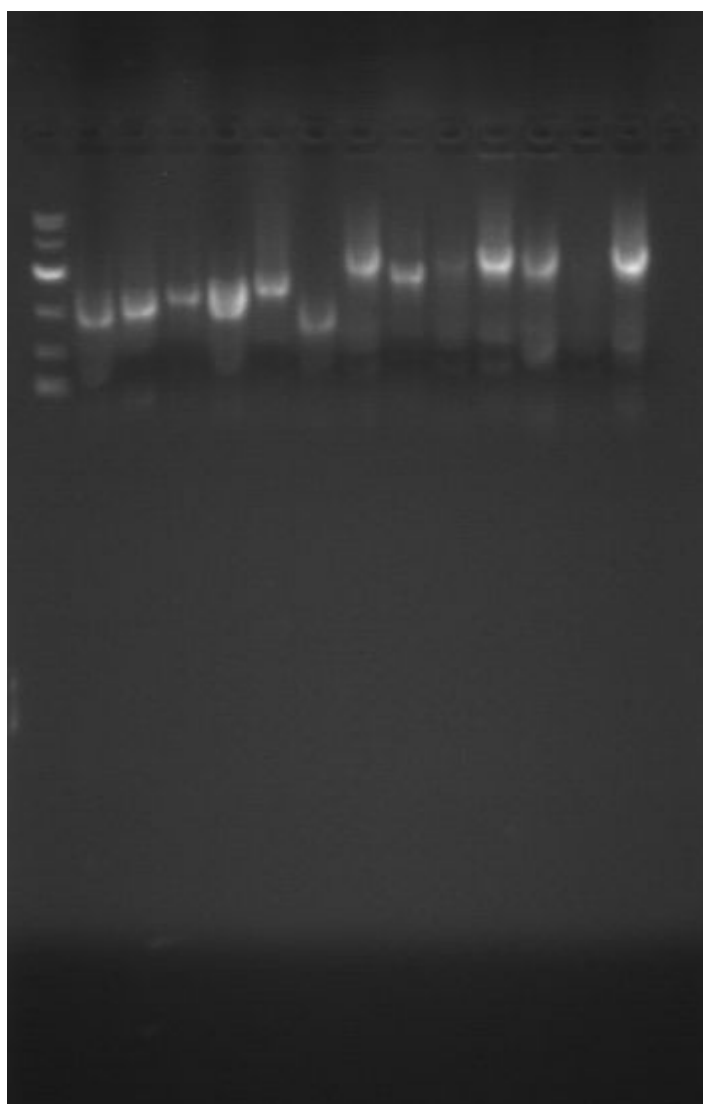


Figure: From left to right, Marker (DL10000), Pcons2, Pcons2-rbs-luxR, Pcons2-rbs-luxR-Ter, Pcons2-rbs-lacI, Pcons2-rbs-lacI-Ter, Prlux-rbs-rfp, Prlux-rbs-rfp-rbs-luxR-Ter, Prlux-rbs-rfp-rbs-luxR, Prlux-rbs-rfp-rbs-luxI-rbs-luxR, Prlux-rbs-rfp-rbs-luxI-rbs-luxR, Prlux-rbs-rfp-rbs-luxI-rbs-luxR-Ter, Prlux-rbs-rfp-rbs-luxI-rbs-luxR-Ter.

G.

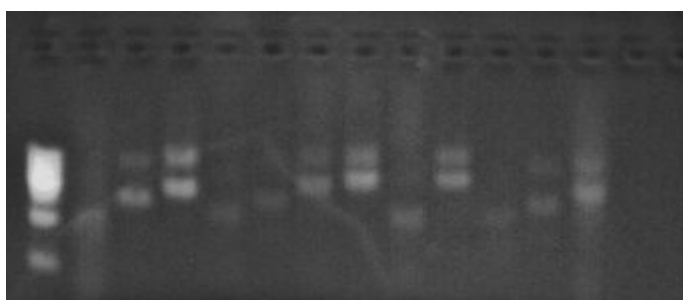
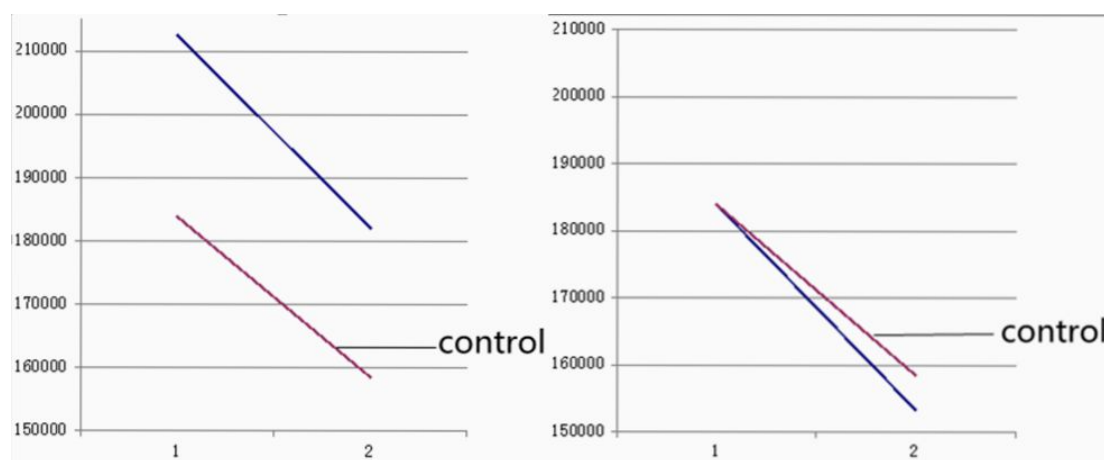


Figure: From left to right, Marker(1Kb), Pcons-rbs-luxR-Ter, Pcons-rbs-luxR-Ter-Prlux-rbs-lacI-Ter, Pcons-rbs-luxR-Ter-Prlux-rbs-lacI-Ter-Plac-rbs-rfp-Ter, Pcons-rbs-lacI-Ter, Pcons-rbs-lacI-Ter-Plac-rbs-luxI-Ter, Pcons-rbs-lacI-Ter-Plac-rbs-luxI-Ter-Pcons-rbs-luxR-Ter, Pcons-rbs-lacI-Ter-Plac-rbs-luxI-Ter-Pcons-rbs-luxR-Ter-Prlux-rbs-rfp-Ter, Pcons-rbs-lacI-Ter-Plac-rbs-luxI-Ter-Pcons-rbs-luxR-Ter-Prlux-rbs-rfp-rbs-luxI-Ter, Pcons-rbs-lacI-Ter-Plac-rbs-luxI-Ter-Pcons-rbs-luxR-Ter-Prlux-rbs-rfp-rbs-luxI-rbs-luxR-Ter, PgolTS-golB-PgolB-rbs-tetR-Ter, PgolTS-golB-PgolB-rbs-tetR-Ter-Ptet-rbs-rfp-Ter, PgolTS-golB-PgolB-rbs-tetR-Ter-Ptet-rbs-rfp-Ter-Pcons-rbs-lacI-Ter.

Part 2:HPLC



The curve above represents our bacteria with plasmid.(left)

To compare the slope of the curves, we move the one above(right)

From the HPLC result, we can conclude that express MlrD in bacteria can increase their MC absorbing ability. Although the concentrations of microcystin in two treatments at 0h

are different, the decrease of MC in the first hour is faster in MlrD bacteria than it in the control (bacteria without MlrD). The difference at 0h can be explained by the fact that IPTG can kill some of the bacteria, which makes the number of bacteria in the IPTG (+) tube is less than the number in the control tube, and that can result in the absorbing quantity of MlrD bacteria lower than control one at the beginning (So the concentration of MC in this tube is higher at 0h). This fact can also explain the phenomenon that there is a decrease of MC concentration in the control group during the one hour cultivation.